The Effects of Some Seed Priming Treatments on Germination and Seedling Development in Wheat

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Abstract

Drought stress has negative effects on plant metabolism and growth. Seed-priming is one of the ways used to strengthen crop plants at the seed stage against drought stress. In this study, the physiological effects of seed-priming (1, 12, 24, and 48 hours) and foliar treatments of H_2O_2 (50 μ M) and GR24 (20 μ M) on 21-day-old wheat seedlings (drought-sensitive *Triticum aestivum* cv. 95 and drought-tolerant cv. Tosunbey) were studied. The plant samples were used to determine the total chlorophyll content, total protein amount, root and stem length, seedling weight, and specific leaf area. Our results showed that only GR24 increased growth in the Tosunbey variety with seed priming. On the other hand, foliar GR24 was effective in both varieties against the detrimental effect of PEG-induced osmotic stress, while foliar H_2O_2 treatment was effective only in the Tosunbey variety. Generally, it has been determined that the GR24 application supports the growth in both varieties, while the H_2O_2 treatment increases the growth only in drought-tolerant wheat variety.

Keywords: Wheat, Drought stress, Seed-priming, GR24, H₂O₂, germination, growth.

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1. Introduction

Abiotic (drought, salinity, temperature, etc.) and biotic (viruses, parasites, etc.) stress factors reduce the crop yield in agriculture by limiting the growth and development of plants (Kaur et al., 2008; Baky et al., 2016). However, the stress tolerance of plants varies depending on the severity of the stress. Stress-tolerant plants that can cope with the increase in stress can survive, while stress-sensitive plants can die. Drought stress has negative effects on plant metabolism and growth and causes serious yield losses in grain production in the world. One of the first responses of plants under water scarcity is to close their stomata to conserve water. In this condition, disruption of gas balance and photosynthetic electron flow results in an imbalance in photosynthesis. One of the earliest signals of drought in plants is the formation of Reactive Oxygen Species (ROS) that alter enzyme activities (Laxa et al., 2019). However, excessive ROS accumulation due to insufficient antioxidant defense capacity causes oxidative stress in plants (Apel and Hirt 2004, Abid et al., 2018, Hasanuzzaman et al., 2018).

ROS concentration increases in plant cells under mild and/or severe stress conditions and these can disrupt metabolism by damaging lipids, proteins, and DNA in the cell. Plant cells are protected from these harmful effects of ROS with the help of antioxidant defense system (AOS) (Asada, 2006). It consists of enzymatic (APX, Ascorbate peroxidase; CAT, Catalase; SOD, Superoxide dismutase; GR, Glutathione reductase) and non-enzymatic antioxidants (tocopherols, carotenoids, water-soluble reductants) (Fazeli et al., 2007). The activity of AOS in stress tolerance is regarded as a selective criterion for reducing yield loss due to stress, especially in the production of agricultural plants.

Wheat is a strategic culture plant produced for food in many countries of the world (FAO 2018). Since it is rich in mineral substances, vitamin B and micronutrients, it meets most of the nutrient and energy needs of the global population (Cummins and Thomson, 2009). Drought conditions drastically reduce photosynthesis, metabolic processes, biomass, and yield in wheat. On the other hand, drought stress responses in wheat vary at morphological, physiological, molecular, and biochemical levels (Liu et al., 2019). It has been shown that inhibition caused by drought in the amount of chlorophyll, relative water content, shoot and root length in durum wheat decreases with the addition of foliar Silicon (Othmani et al., 2020). In addition, it was determined that the rosehip extract treatment in two wheat varieties with different tolerance to drought increased the antioxidant capacity through different antioxidant enzymes (Baltacier, 2019).

To prevent the decrease in product yield caused by stress factors, new stress-resistant varieties are being developed. In this context, priming treatments include some preliminary applications to reveal the stress tolerance potential of existing varieties. Currently, various priming techniques are applied from farm to industrial level for better crop yield (Figure 1, Haider et al., 2019). Hydro-priming revitalizes seeds soaked in sterile water at the appropriate temperature and ensures a robust crop formation, especially in tropical regions. Osmo-priming provides control of radicle growth due to water absorption in seeds incubated in a salt and polyethylene glycol (PEG) solution. Thus, the seeds protect from low external water potential and synchronized germination. Chemical priming and biopriming are used specifically to provide nutrients to seeds and protect them from any disease attack (Saboor et al., 2019). Seed priming improves drought stress tolerance by increasing the membrane stability, improving plant water condition and chlorophyll content, enhancing germination under normal and stressful conditions. It causes early and uniform germination due to the increased synthesis of metabolites related to germination. These applications can be carried out both at the seed and seedling stage (Saboor et al., 2019). It has been reported that the treatments of salicylic acid (SA) and thiourea (TU) to wheat increase the grain yield under water scarcity conditions (Yadav et al., 2020). On the other hand, drought priming with PEG 3000 (20%) increases the drought tolerance of wheat, but this requires abscisic acid (ABA) and jasmonic acid (JA) (Wang et al., 2021). In addition, drought priming in the early growing stage of wheat increases drought tolerance by removing ABA and H₂O₂ (Wang et al., 2020). Finally, the treatment of polyamines (PAs) as seed priming or foliar treatment to wheat seedlings decreases lipid peroxidation and ROS concentration and increases CAT activity under drought stress. Thus, PAs have been shown to improve chloroplast content and membrane stability in wheat (Hassan et al., 2020).

Strigolactones (SL), a phytohormone, have important functions around the root, including drought tolerance. On the other hand, they promote the symbiotic relationship between plants and soil microorganisms. In addition, they provide the germination of parasitic plant seeds (Kohlen et al.2011). SL treatment has been shown to increase plant resistance under drought conditions (Min et al., 2019; Sedaghat et al., 2020). It has been determined that SL pre-treatments in wheat and sand lily have a promoting role in salt tolerance by stimulating AOS against salt stress (Özel, 2018; Önay, 2019). It has also been reported that the treatment of GR24, a synthetic SL, has a protective effect on plant cells against drought stress (Min et al., 2019).

Hydrogen peroxide is produced in many organelles in the plant cell during photosynthetic electron transport chain and respiration (Foyer et al.1997). Therefore, high H_2O_2 accumulation are lethal to plant cells due to its toxic nature. However, it has also been shown that H_2O_2 in non-toxic concentrations can act as a signal molecule in protecting plants against biotic and abiotic stress (Caverzan et al.2016; Küçükkarakaş, 2017; Arıcan, 2019). H_2O_2 can spread across the cell membrane and be transported to other compartments where they can function as signal molecules or be destroyed (Caverzan et al.2016). On the other hand, it has also been shown that H_2O_2 pretreatment can be effective in reducing the harmful effects of drought stress in wheat and rice (Shan et al., 2018; Sohag et al., 2020).

In this study, the physiological effects of H_2O_2 , GR24 (synthetic SL), and PEG 3000 (%20) on seed germination and seedling development in two wheat varieties (drought sensitive-*Triticum aestivum* cv. Sultan-95 and drought tolerant-cv. Tosunbey) were investigated.



Figure 1. Priming Types (Haider et al., 2019)

2. Materials and Methods

2.1. Plant Material

In this study drought sensitive-cv. Sultan-95 and drought tolerant-cv. Tosunbey varieties of *Triticum aestivum* L. (*Poaceae*) were used. The seeds of Sultan-95 were obtained from Eskişehir Geçit Kuşağı Agricultural Research Institute and the seeds of Tosunbey from Ankara Field Crops Central Research Institute. Wheat seeds were sterilized by soaking in 5% sodium hypochlorite solution for 5 minutes and washing with sterile distilled water. The seeds germinated by keeping them in moist filter papers for 3 days were then transferred to petri dishes and pots containing perlite. Wheat seedlings were grown in the plant growth cabinet at 22-24 $^{\circ}$ C in the 16:8 h light/dark photoperiod. Wheat seedlings in pots were irrigated with Hoagland nutrient solution (100%) for 21 days (Steward, 1983).

2.2. Priming Treatments

 H_2O_2 (50 μ M) and GR24 (20 μ M) priming treatments were treatment to sterile wheat seeds in petri dishes for 1, 12, 24 and 48 hours. After the seeds were washed with distilled water, they were planted on petri dishes moistened with distilled water and their germination was followed for 7 days. At the end of the experiment, the seedlings were harvested, and these samples were used to determine the total chlorophyll content, total protein amount, root and shoot length, seedling weight (biomass), germination (%) and specific leaf area.

 H_2O_2 (50 µM) and GR24 (20 µM) priming treatments were carried out by leaf spraying on 21-day-old wheat seedlings grown in pots containing perlite. Osmotic stress mediated drought stress was started with the Hoagland solution containing PEG 3000 (20%) at 48 h after other priming treatments. Leaf samples were collected from wheat seedlings on the 7th and 14th days following the drought treatment and were used to determine the total chlorophyll content, total protein amount, root and shoot length, seedling weight and specific leaf area.

2.3. Physiological and Biochemical Parameters

2.3.1. Root-Shoot Length

The lengths of the root and shoot parts of the plants were measured with a ruler.

2.3.2. Chlorophyll Analysis

Total chlorophyll amounts of leaf samples were measured with chlorophyll meter (Minolta, SPAD-502) (Peryea and Kammereck, 1997). Data were made in 15 replicates on different leaves in the plant groups.

2.3.3. Determination of Seedling Weight

Seedling weights were determined by weighing on a precision scale (gr). In the petri experiment, the weight of 10 seedlings from each group was weighed in a way that all repeats from each group were taken.

2.3.4. Determination of Germination Percentage

At the end of the experiment, it was determined by the ratio of germinated seed to the total number of seeds in petri dishes (%) (Zhang et al., 2020).

2.3.5. Determination of Specific Leaf Area (SLA)

Specific leaf area was calculated using the leaf photos of wheat seedlings in the Image J program. Then the samples are dried in an oven at 70° C for 24 h and weighed on a precision scale. SLA is calculated by the formula (Wilson et al. 1999): SLA = Area (cm^2) / Dry weight (mg^{-1})

2.3.6. Total Protein Amount

Tissue samples were homogenized with 50 mM Na-P (pH: 7.8 and 1 mM EDTA) buffer and then centrifuged at 10000 rpm. The supernatants obtained after centrifugation are mixed with protein reagent containing 0.1 g of Coomassie Brilliant Blue G 250, 50 mL of ethanol and 100 mL of orthophosphoric acid in the tube. Sample protein content is obtained by calculating the absorbance at 595 nm in the spectrophotometer (mg / mL) in the protein standard graph (Bradford, 1976).

2.3.7. Statistical analysis

The data were made with Tukey test using one-way analysis of variance (ANOVA). SPSS (Statistical Package for the Social Sciences, version 20.0) program was used for statistical analysis.

3. Results and Discussion

3.1 Petri Experiment

3.1.1. Root-Shoot Length

Root length increased 38% with GR24 in Tosunbey with 1 h seed priming treatment compared to control. In Sultan-95, the increases with GR24 and H_2O_2 treatments were determined as 19% and 25%, respectively. While the shoot length increased by 12% in Tosunbey variety with H_2O_2 treatment, a decrease of 4.5% and 9% was determined with GR24 and H_2O_2 treatment, respectively, in Sultan-95 variety compared to the control (Figure 2a).

The 12 h seed priming treatment increased the root length in both varieties compared to the control. These increases are 19.5% and 24%, respectively, in the Tosunbey variety with GR24 and H_2O_2 treatments, while in the Sultan-95 variety it is 33% and 18%. Shoot length increased 16% with H_2O_2 treatment in Tosunbey variety compared to control, while it decreased by 19% in Sultan-95 compared to control with H_2O_2 treatment (Figure 2b).

24 h GR24 and H_2O_2 treatments increased root length by 19% and 14%, respectively, in Tosunbey variety compared to control. Interestingly, root length increased 1.7 times higher than the control with GR24 treatment in Sultan-95 variety. In this variety, H_2O_2 treatment increased the root length by 22%. Shoot length has increased with both treatments in Tosunbey variety. These increases were determined as 25% and 20% for GR24 and H_2O_2 treatments, respectively. In Sultan-95 variety, shoot length increased by 9.5% with the treatment of GR24 and decreased by 8% with H_2O_2 (Figure 2c).

While 48 h GR24 treatment decreased the root length by 16% in Tosunbey cultivar, it increased 9.7% with H_2O_2 treatment. In Sultan-95 variety, root length increased with both treatments. These increases were determined as 34% and 53%, respectively, for GR24 and H_2O_2 treatments compared to the control. Shoot length is increased in both varieties compared to the control in all treatments. These increases were found 11% and 1.7 times for GR24 and H_2O_2 treatments in Sultan-95, respectively (Figure 2d).

3.1.2. Total Chlorophyll Content

Total chlorophyll content increased by 1-4% in both treatment groups with 1, 12, 24 h treatments, while GR24 and H_2O_2 pre-treatments decreased by 14% and 9%, respectively, in 48 h of treatment in Tosunbey (Figure 3a).

In Sultan-95 variety, total chlorophyll content decreased with 12, 24 and 48 h treatments compared to the control. These reductions were 37% and 25% for 12 h GR24 and H_2O_2 priming, and 21% and 13% for 24 h, respectively. 1 h of GR24 and H_2O_2 treatment did not change the amounts of chlorophyll (Figure 3b).



Figure 2. Effects of GR24 and H₂O₂ seed priming on root and shoot length in 7d wheat seedlings. (a: 1 h, b: 12 h, c: 24 h, d: 48 h, TK: Tosunbey control, TGR24: Tosunbey GR24, TH₂O₂: Tosunbey H₂O₂, SK: Sultan-95 control, SGR24: Sultan-95 GR24, SH₂O₂: Sultan-95 H₂O₂).

3.1.3. Seedling Weight (Biomass)

In Tosunbey variety, seedling weight increased with 1 h and 12 h priming in both treatment groups. These increases are 15% and 13%, respectively, in the GR24 group compared to the control. In the 24 h treatment of this variety, a 10% decrease was detected in the GR24 group compared to the control, while an increase of 7.5% was found in the H_2O_2 group. In 48 h treatments, it was followed by a 9.5% decrease in the GR24 group and a 10% increase in the H_2O_2 group. As a result, H_2O_2 priming increased biomass in all treatments compared to control, while GR24 treatment increased with only 1 h and 12 h priming (Figure 4a).

In Sultan-95 variety, 1 h and 12 h H_2O_2 priming reduced biomass by 18% and 10%, respectively, compared to control. Conversely, seedling weight increased by 25% and 16% in 24 h treatments of GR24 and H_2O_2 , while it was 12% and 16% in 48 h treatments, respectively. As a result, 1 h and 12 h treatments of this variety decreased biomass, while 24 h and 48 h treatments increased (Figure 4b).



Figure 3. Effects of GR24 and H₂O₂ seed priming on total chlorophyll content in 7d wheat seedlings, a: Total chlorophyll content of Tosunbey variety, b: Total chlorophyll content of Sultan-95 variety (1 h, 12 h, 24 h, 48 h: priming times, K: Control, GR24: GR24 treatment, H₂O₂: H₂O₂ treatment).



Figure 4. Effects of GR24 and H₂O₂ seed priming on seedling weight (biomass) in 7d wheat seedlings, a: cv. Tosunbey, b: cv. Sultan-95 (1 h, 12 h, 24 h, 48 h; priming times; K: Control, GR24: GR24 treatment, H₂O₂: H₂O₂ treatment).

3.1.4. Germination Percentage (%)

In Tosunbey cultivar, all seeds in both groups were germinated in 1 h priming. On the other hand, germination decreased in all treatments of 12, 24 and 48 h. While these decreases were 19% in 12 h H_2O_2 treatment compared to control, it was determined as 14% in 24 h GR24 treatment (Figure 5a). In Sultan-95 variety, 1 h treatment decreased the germination percentage by 4% and 14% compared to the control for GR24 and H_2O_2 priming, respectively. The 12 h treatment decreased the germination by 10% in both priming treatments. Interestingly, while 24 h treatment of GR24 decreased germination by 4%, all seeds were germinated with the 12 h treatment decreased the germination by 10% in both priming treatments. Germination decreased by 19% with GR24 treatment in 48 h treatments (Figure 5b).



Figure 5. Effects of GR24 and H_2O_2 seed priming on germination in 7d wheat seedlings, a: cv. Tosunbey, b: cv. Sultan-95 (1 h, 12 h, 24 h, 48 h; priming times; K: Control, GR24: GR24 treatment, H_2O_2 : H_2O_2 treatment).

3.1.5. Specific Leaf Area (SLA)

Specific leaf area increased by 30% and 20%, respectively, with 1 h and 24 h GR24 priming in Tosunbey compared to the control and decreased with the other 12 h and 24 h treatments. H_2O_2 application has increased SLA in all treatments compared to control in this type. These increases were determined as 40%, 14%, 13% and 9% for 1, 12, 24, and 48 h, respectively (Figure 6a). SLA decreased in Sultan-95 variety, except for 48 h H₂O₂ priming. Accordingly, while SLA decreased by 19-30% with GR24 priming treatments, it was found to decrease 7-45% with H₂O₂ priming treatments (Figure 6b).

3.1.6. Total Protein Amount

In Tosunbey variety, 1, 12 and 24 h GR24 and H_2O_2 treatments significantly increased the total protein amounts compared to the control. These increases were determined as the lowest 11% (12 h) and the highest 45% (24 h) for GR24, while the lowest 15% (12 h) and the highest 39% (24 h) for H_2O_2 (Figure 7a).

In Sultan-95 variety, GR24 treatments significantly increased the total protein amount by 14% compared to the control with only 12 h application. Other GR24 applications did not change the total protein content. All applications except 12 h of H_2O_2 application significantly increased the total protein content of this variety by 11-30% compared to the control. The highest increase was observed in 24 h application (Figure 7b).



Figure 6. Effects of GR24 and H₂O₂ seed priming on SLA in 7d wheat seedlings, a: cv. Tosunbey, b: cv. Sultan-95 (1 h, 12 h, 24 h, 48 h; priming times; K: Control, GR24: GR24 treatment, H₂O₂: H₂O₂ treatment).



Figure 7. Effects of GR24 and H₂O₂ seed priming on total protein amounts in 7d wheat seedlings, a: cv. Tosunbey, b: cv. Sultan-95 (1 h, 12 h, 24 h, 48 h; priming times; K: Control, GR24: GR24 treatment, H₂O₂: H₂O₂ treatment).

3.2. Pot Experiment 3.2.1. Root-Shoot Length

In the control group [PEG (-)] shoot length decreased by 3% with GR24 treatment on the 7th day compared to the control in Tosunbey variety, while it increased by 1.2% with H_2O_2 treatment. In Sultan-95 variety, shoot length decreased by 4-6% with all treatments. Root length increased by 13% on the 7th day with GR24 treatment in Tosunbey variety and decreased by 20% with H_2O_2 treatment. In Sultan-95 variety, root length increased by 13% on the 7th day with GR24 treatment. Shoot length of Tosunbey variety decreased by 7% compared to control with H_2O_2 treatment. Shoot length of Tosunbey variety decreased by 7% compared to control with H_2O_2 treatment on the 14th day. In Sultan-95, shoot length increased by 7% with GR24 priming and 4% with H_2O_2 priming compared to the control. Root length increased by 9% with GR24 and H_2O_2 on the 14th day in Tosunbey variety. In Sultan-95 variety, root length increased by 7% on the 14th day with all treatments.

At the end of the experiment, it was determined that drought treatment [PEG (+)] decreased shoot length in all priming treatments in both varieties. While these decreases are 6-10% on the 7th and 14th days in Tosunbey variety, it is 10-16% on the 7th day in Sultan 95 variety and 1-4% at the end of the trial. It was determined that the root length did not show a significant change on the 7th day with priming treatments in Tosunbey and decreased by 8% with 4% H_2O_2 treatment on the 14th day with GR24 treatment. In Sultan-95 variety, GR24 treatment decreased the root length by 21-24% and H_2O_2 treatment by 11-15% during the trial (Figure 8).



Figure 8. Effects of foliar GR24 and H₂O₂ treatments on root and shoot lengths in 7d- and14d-old wheat seedlings under control [PEG (-)] and drought treatment [PEG (+)] conditions (TK: Tosunbey control, TGR24: Tosunbey GR24, TH₂O₂: Tosunbey H₂O₂, SK: Sultan-95 control, SGR24: Sultan-95 GR24, SH₂O₂: Sultan-95 H₂O₂).

3.2.2. Total Chlorophyll Amount

The amount of total chlorophyll increased with GR24 priming at the end of the experiment under control and drought conditions in both varieties. These increases were 15% in Tosunbey variety in PEG (-) group on the 14th day and 7% in PEG (+) group. In Sultan-95 variety, these increases are 7% and 10%, respectively (Figure 9a-b). In contrast, H_2O_2 priming decreased the total chlorophyll content at the end of the experiment in both varieties under control and drought conditions. While these decreases were 2% in Tosunbey variety in PEG (-) group on the 14th day, it was 7% in PEG (+) group. In Sultan-95 variety, these decreases are 8% and 7%, respectively (Figure 9c-d).

3.2.3. Seedling Weight (Biomass)

GR24 priming increased biomass in the PEG (-) group on the 7th and 14th day, respectively, by 14% and 20% compared to the control in Tosunbey variety. On the contrary, drought treatment decreased biomass by 27% and 14%, respectively, compared to control. In the GR24 priming Sultan 95, the biomass was decreased at the end of the experiment under control and drought conditions. These decreases were 36% and 52%, respectively, in the PEG (-) group on the 7th and 14th days, and 6% and 13% in the PEG (+) group, respectively (Figure 10a-b).

 H_2O_2 priming reduced biomass at the end of the experiment under control and drought conditions in both varieties, except for the 7th day of Tosunbey variety. At the end of the experiment, these decreases were determined as 18% in Tosunbey in PEG (-) group and 8% in PEG (+) group. In Sultan-95 variety, these decreases were 52% and 57%, respectively, on the 7th day, and 17% and 10% at the end of the experiment (Figure 10c-d).

3.2.4. Specific Leaf Area (SLA)

It was determined that SLA decreased by 5% and 3% on the 7th and 14th day of the experiment under control and drought conditions with GR24 priming in Tosunbey variety. It was determined that there was no significant change in the control group due to GR24, and in the drought group, SLA increased by 9% during the experiment in Sultan-95 (Figure 11a-b).

Interestingly, H_2O_2 priming increased SLA by 11% in the control group of Tosunbey during the experiment. However, the drought treatment in this variety has not changed the SLA. In Sultan-95, H_2O_2 priming decreased SLA by 4% on the 7th day in the control group and increased it by 4% at the end of the experiment. The drought treatment decreased the SLA of this variety by 8% and 3% on the 7th and 14th days, respectively (Figure 11c-d).



Figure 9. Effects of foliar GR24 and H₂O₂ treatments on total chlorophyll contents in 7d- and14d-old wheat seedlings under control [PEG (-)] and drought treatment [PEG (+)] conditions (TK: Tosunbey control, TGR24: Tosunbey GR24, TH₂O₂: Tosunbey H₂O₂, SK: Sultan-95 control, SGR24: Sultan-95 GR24, SH₂O₂: Sultan-95 H₂O₂).

3.2.5. Total Protein Content

While the total protein content of the Tosunbey variety increased by 23% and 108%, respectively, on the 7th and 14th days of the control group with GR24 priming, it increased by 49% and 1% in the drought group, respectively. In this cultivar, the amount of protein increased by 63% and 50% on the 7th and 14th days in the control group with H_2O_2 priming, respectively. However, in the drought group, it increased by 51% on the 7th day and decreased by 18% on the 14th day (Figure 12a-b).

In the Sultan-95 variety, the amount of protein increased by 11% and 1%, respectively, on the 7th and 14th days of the control group with GR24 priming, while it decreased by 8% on the 7th day in the drought group and increased by 39% on the 14th day. In this cultivar, the amount of protein with H_2O_2 priming decreased by 2% and 45% on 7th and 14th days in the control group, respectively. However, in the drought group, it increased by 35% and 14% on days 7 and 14, respectively (Figure 12c-d)



Figure 10. Effects of foliar GR24 and H₂O₂ treatments on seedling weight in 7d- and14d-old wheat seedlings under control [PEG (-)] and drought treatment [PEG (+)] conditions (TK: Tosunbey control, TGR24: Tosunbey GR24, TH₂O₂: Tosunbey H₂O₂, SK: Sultan-95 control, SGR24: Sultan-95 GR24, SH₂O₂: Sultan-95 H₂O₂).



Figure 11. Effects of foliar GR24 and H₂O₂ treatments on SLA in 7d- and14d-old wheat seedlings under control [PEG (-)] and drought treatment [PEG (+)] conditions (TK: Tosunbey control, TGR24: Tosunbey GR24, TH₂O₂: Tosunbey H₂O₂, SK: Sultan-95 control, SGR24: Sultan-95 GR24, SH₂O₂: Sultan-95 H₂O₂).





Figure 12. Effects of foliar GR24 and H₂O₂ treatments on SLA in 7d- and14d-old wheat seedlings under control [PEG (-)] and drought treatment [PEG (+)] conditions (TK: Tosunbey control, TGR24: Tosunbey GR24, TH₂O₂: Tosunbey H₂O₂, SK: Sultan-95 control, SGR24: Sultan-95 GR24, SH₂O₂: Sultan-95 H₂O₂).

3.3. Discussion

In the last decade, there have been numerous reports of newly discovered carotenoid derivative strigolactones (SLs) reducing the negative effects of stress on plants under drought stress (Min et al., 2019; Omoarelojie et al., 2020; Sedaghat et al., 2017; Ni et al., 2020; Lin et al., 2020). Accordingly, foliar application of GR24 can reduce the negative effects of drought stress by increasing the amount of chlorophyll in the grape (Min et al., 2019), can increase seed germination and seedling development in lupine (Omoarelojie et al., 2020), and can also increase tolerance to drought stress in wheat (Sedeghat et al., 2017). In addition, the application of GR24 (0,18 μ ML⁻¹) to colza seed can significantly increase lateral root growth, shoot growth, and root/shoot ratio in seedlings. Furthermore, it has been shown that exogenous SL application can alleviate the negative impact of drought stress on the growth of colza seed seedlings (Lin et al., 2020).

Similarly, our results showed that the application of GR24 priming in drought-resistant cv. Tosunbey and drought-sensitive cv. Sultan-95 increases root lengths. The highest increase was obtained with 1 h GR24 application in Tosunbey variety and 24 h in Sultan-95 variety. Increased the shoot length in these varieties of 24 h GR24 application. In the pot trial, it was determined that all GR24 applications in the control group increased the root length. Contrary to other studies, our results showed that exogenous GR24 decreased root and shoot lengths in wheat under PEG-induced osmotic stress. Exogenous GR24 has been shown to decrease the endogenous auxin level in rapeseed by 52% and 75% 12 hours and 1 day after administration (Ni et al., 2020). This may be explanatory for the increase in root length of control plants in the pot trial. However, more detailed research is needed for growth inhibition in seedlings under drought.

It was determined that all GR24 applications in Tosunbey increased the amount of chlorophyll in both petri and pot experiments. In the drought-sensitive variety Sultan-95, it was determined that the amount of chlorophyll decreased in the petri and increased in the pot trial. These results are consistent with those in grape (Min et al., 2019), lupine (Omoarelojie et al., 2020), and rapeseed (Lin et al., 2020).

On the other hand, a decrease in biomass has been reported in wheat seedlings under drought stress (Shan et al., 2018). 1 h and 12 h GR24 priming applications in Tosunbey, 24 h, and 48 h GR24 priming applications in Sultan-95 increased the biomass. Interestingly, at the end of the pot trial, biomass decreased due to osmotic stress-dependent drought. According to the biomass results, GR24 treatment improved the negative effects of drought in wheat varieties exposed to drought stress. Moreover, GR24 priming treatments did not increase the germination percentage.

Specific leaf area (SLA) is the ratio of leaf biomass to leaf area and is a measure of plant strategy in response to environmental changes (Zhou et al., 2020). Our study results showed that SLA increased with GR24 priming and only with GR24 in Sultan-95 and Tosunbey seedlings under drought stress. While the total protein content increased with all GR24 priming applications in Tosunbey variety, it increased with only 12 h GR24 in Sultan-95. In the pot trial, it was determined that the total amount of protein increased on the 7th day and at the end of the experiment, respectively, in Tosunbey and Sultan-95 seedlings under drought stress with foliar GR24 treatment. These results are consistent with those for data of rapeseed under drought stress (Lin et al., 2020).

Hydrogen peroxide (H_2O_2) is normally a toxic molecule to the cell, however, at low concentrations acts as a signal molecule. It was showed that priming treatment or exogenous treatment of H_2O_2 in plants exposed to drought stress improves the harmful effects of stress. (Shang et al., 2020; Iqbal et al, 2018; Habib et al., 2020). Although our results show that root lengths increased in both varieties with H_2O_2 priming, it was determined that shoot lengths increased only in Tosunbey effectively. These results are consistent with the results from mung bean seeds under drought stress (Nawaz et al., 2021). Pot experiment results indicated that increases in root and shoot lengths were inhibited by osmotic stress caused by PEG. These results are not compatible with H_2O_2 priming results obtained in rice (Shang et al., 2020), quinoa (Iqbal et al., 2018), and wheat (Habib et al., 2020) under drought stress with H_2O_2 . Additionally, H_2O_2 priming increased the amount of chlorophyll in Tosunbey but foliar H_2O_2 treatment decreased it. Further, seed germination increased with 24 h H_2O_2 priming in Sultan-95.

These results indicate that the drought-tolerant variety shows better growth than the sensitive variety. In fact, all H_2O_2 priming treatments increased biomass in Tosunbey, while Sultan-95 is limited to 24h and 48 h. On the other hand, it was determined that osmotic stress decreased with foliar H_2O_2 treatments in the pot experiment. In addition, H_2O_2 priming and foliar H_2O_2 treatments appear to be effective in increasing the SLA of Tosunbey variety. In this context, it was found that the total protein content increased in both varieties with H_2O_2 priming, with foliar H_2O_2 treatment increased it only in the drought group.

Based on our results, it can be said growth parameters and total protein to remedy the harmful effect of PEG-induced osmotic stress showed that foliar GR24 is effective in both wheat varieties, while foliar H_2O_2 is effective only in the Tosunbey variety. Accordingly, our results indicate that GR24 priming promotes growth in both varieties, while H_2O_2 priming increases growth in Tosunbey varieties.

4. Conclusion

In this study, results were obtained that GR24 applications, a synthetic SL, support growth and yield in drought-resistant Tosunbey variety. The failure to achieve similar results in H_2O_2 applications may be due to the phytohormonal nature of GR24 or the concentration of H_2O_2 . For this reason, new research is needed on the relationship between GR24 and other phytohormones and on the different concentrations of H_2O_2 .

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